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(51) International Patent Classification 7: (11) International Publication Number: WO 00/27996 C12N 5/00 A1 (43) International Publication Date: 18 May 2000 (18.05.00) (81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, (21) International Application Number: PCT/EP99/08482 (22) International Filing Date: 8 November 1999 (08.11.99) ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, (30) Priority Data: SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, 60/107,646 9 November 1998 (09.11.98) US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, US LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, (71) Applicants (for all designated States except US): CONSORZIO PER LA GESTIONE DEL CENTRO DI BIOTECNOLO-MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GIE AVANZATE [IT/IT]; Largo Rosanna Benzi, 10, GA, GN, GW, ML, MR, NE, SN, TD, TG). I-16132 Genova (IT). ISTITUTO NAZIONALE PER LA RICERCA SUL CANCRO [IT/IT]; Largo Rosanna Benzi, Published 10, I-16132 Genova (IT). With international search report. (72) Inventors; and Before the expiration of the time limit for amending the (75) Inventors/Applicants (for US only): CANCEDDA, Ranieri claims and to be republished in the event of the receipt of [IT/IT]; Via Nizza, 11, I-16145 Genova (IT). DOZIN, amendments. Beatrice [IT/IT]; Via S. Massimo, 111, I-16035 Rapallo (74) Agent: MINOJA, Fabrizio; Bianchetti Bracco Minoja S.r.l., Via Rossini, 8, I-20122 Milan (IT). (54) Title: SERUM FREE MEDIUM FOR CHONDROCYTE-LIKE CELLS (57) Abstract The present invention provides serum free media for growth and proliferation of chondrocytes and mesenchymal stem cells in culture.

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SERUM FREE MEDIUM FOR CHONDROCYTE-LIKE CELLS

BACKGROUND OF THE INVENTION

Bone and cartilage transplantation is an absolute need in reconstruction of bone and cartilage segments in plastic surgery, traumatic surgery or after the removal of neoplastic lesions, etc. Typically, material of human (autologous, from donors or from cadavers) or animal origin has been used for this purpose. Given the increased demand from clinicians for transplant tissues, the increased need for microbial safety in tissue transplantation, the advances in cell biology, cell differentiation and tissue engineering, the concept of rebuilding tissues from autologous or allogeneic cells expanded in vitro has become a growing field in the world of biomedical sciences. Cellular sources for skeletal repair include chondrocytes and cells committed to the chondrocyte lineage, and mesenchymal stem cells, the former specific for cartilage, the latter multipotential and therefore having the potential to be used to replace bone, cartilage and other tissues.

Mesenchymal stem cells (MSCs) are found in bone marrow as well as in blood, dermis and periosteum. Although these cells are normally present at very low frequencies in bone marrow, these cells can be isolated purified and culturally expanded, for example, as described in U.S. Patent No. 5,486,359.

Typically, the *in vitro* expansion of chondrocytes and MSCs takes place in culture medium supplemented with bovine serum or optimally with autologous serum from the patient. However, the presence of animal or autologous serum in chondrocyte and MSC cultures has certain disadvantages and limitations in view of the potential therapeutical applications of these cultures.

For example, serum is not the physiological fluid most cells closely contact in tissue in vivo. This is particularly true for chondrocytes that, in

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vivo, are embedded in their avascularized matrix and rely for their own growth and differentiation on various growth factors and cytokines acting in an autocrine/panacrine manner rather than diffusing from the distant bloodstream. Further, there is often high variability between animal serum batches. Extensive serum screening required to select the batch most representative of the *in vivo* inductive effects can be time-consuming and expensive The preparation of autologous serum from patients is also time consuming and supplies are limited. Animal serum can further potentially carry unknown pathogens with consequent risk of contamination for the patient.

Thus, serum substitutes for culturing cells for potential in vivo therapeutic applications is desirable.

SUMMARY OF THE INVENTION

The present invention provides a serum substitute for culturing cells in vitro using well defined factors able to support cell viability, proliferation and differentiation as effectively as serum containing medium. In a preferred embodiment, the cells are articular chondrocytes or mesenchymal stem cells.

In one aspect, the invention comprises a composition for the expansion of chondrocytes, comprising a minimum essential medium, a growth factor, albumin, a steroid, an antioxidant, an iron source, a fatty acid and/or a lipid source, and insulin. In a particularly preferred embodiment, the serum free growth medium for chondrocytes comprises FGF-2 as a growth factor, linoleic acid as the lipid/fatty acid source, ascorbic acid and β -mercaptoethanol as antioxidants, holo- and apo-transferrin as the iron source, and dexamethasone as a steroid. Optional ingredients can include cholesterol, trace metals such as selenium, and vitamins such as biotin and sodium pantotenate.

In another aspect, the invention comprises a composition for the maintenance of mesenchymal stem cells, comprising a growth factor, albumin,

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a steroid, an antioxidant, an iron source, a fatty acid and/or a lipid source, one or more vitamins, one or more trace metals, and IGF-1, in combination with a minimum essential medium. In a particularly preferred embodiment, the serum free growth medium for mesenchymal stem cells comprises FGF-2, LIF and SCF as growth factors, sodium pantotenate and biotin as vitamins, and selenium as a trace metal.

BRIEF DESCRIPTION OF THE DRAWING

Figure 1 shows the comparison of articular chondrocyte growth kinetics in medium containing FCS and the serum free medium of the present invention.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides serum free compositions suitable for chondrocyte and mesenchymal stem cell growth and proliferation. The compositions may include in a base minimum essential medium, such as Coon's modified Ham's F-12 medium, the following components as a substitute for serum:

- i) one or more growth factors or proteins which cause resting cells to undergo cell division and/or differentiation, such as insulin, FGF-2, PDGFbb, EGF, LIF and SCF and IGF-1;
 - ii) one or more steroids such as dexamethasone;
- iii) one or more sources of lipids and fatty acids, necessary for cell membrane biosynthesis, such as cholesterol and linoleic acid; and
 - iv) an iron source such as transferrin.
- FGF-2, PDGFbb and EGF are potent mitogens for cells of mesenchymal origin. Dexamethasone is known to keep cells in a cycling phase in vitro.

The serum free medium of the present invention may further comprise:

i) albumin (preferably of mammalian species) which functions as an aspecific carrier;

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- ii) one or more antioxidants such as β-mercaptoethanol;
- iii) a supplement for coenzyme transport in carboxyl group transfer reactions, such as biotin;
- iv) trace elements as a supplemental source of metal necessary for electron transport and many metalloenzymes and proteins, such as selenium;
 - v) vitamins, such as biotin and pantotenate; and
 - vi) ascorbic acid, to facilitate organization of the extracellular matrix.

Insulin and dexamethasone are added at the average concentrations usually reported in the literature.

IGF-I, LIF and SCF are present at concentrations in the range from about 5 to about 10 ng/ml; preferably at a concentration of 5 ng/ml. All the other components are included in a range of concentration typically used in cell culture studies.

In a preferred embodiment of the composition suitable for the growth and proliferation of the chondrocytes, the defined components comprise EGF, PDGFbb and FGF-2, ascorbic acid, linoleic acid, human serum albumin (HSA), β-mercaptoethanol, dexamethasone, insulin, human holo- and apotransferrin. In this embodiment, FGF-2, PDGFbb and EGF are present at concentrations in the range of from about 1 to about 10 ng/ml. In a preferred embodiment, FGF-2, PDGFbb and EGF are present at concentrations of from 1 to 2 ng/ml.

In a preferred embodiment of the composition suitable for the growth and proliferation of the mesenchymal stem cells, the defined components comprise: EGF, PDGFbb and FGF-2, LIF, SCF, IGF-I, ascorbic acid, cholesterol, HSA, β-mercaptoethanol, dexamethasone, human holo- and apotransferrin, selenium, biotin, sodium pantotenate. FGF-2, PDGFbb and EGF are present at concentrations in the range of from about 5 ng/ml to about 10 ng/ml of each factor. The preferred concentrations of FGF-2, PDGFbb and

EGF are 10 ng/ml. FGF-2 alone was found to be the most active factor for maintenance of osteochondrogenic potential in MSCs.

Example 1

Articular cartilage was harvested from the knee joint of young adult human donors. The samples were first cleaned of any adherent muscular, connective or subchondral bone tissues, minced into 1-3mm fragments and rinsed in PBS. Single chondrocytes were then released by repeated enzymatic digestions at 37°C with 0.25% trypsin, 400U/ml collagenase I, 1000U/ml collagenase II and 1 mg/ml hyaluronidase. Trypsin was then blocked and removed by rapid and extensive washes in PBS containing soybean trypsin inhibitor. Cells were plated in anchorage-dependent conditions in Coon's modified Ham's F-12 medium supplemented either with 10% fetal calf serum (FCS, control culture) or the following defined components: EGF, PDGFbb and FGF-2, ascorbic acid, linoleic acid, human serum albumin (HSA), β-mercaptoethanol, dexamethasone, insulin, human holo- and apo-transferrin. Table 1 below shows the preferred amounts of each component. To favor adhesion of the cells in serum free conditions, the dishes were pre-coated with 2% gelatin.

Insulin may not be substituted with IGF-1 in the medium for chondrocytes. Insulin was preferably at a concentration of 5 μ g/ml. Selenium, biotin, sodium pantotenate and cholesterol can be routinely included but are optional.

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Table 1

Medium Supplement for Serum Free Expansion

of Human Chondrocytes

	INGREDIENT	CONCENTRATION
5	(Basal medium: Coon's modified Ha	m's F12)
	FGF-2	1-10 ng/ml
	PDGFbb	1-10 ng/ml
	EGF	1-10 ng/ml
	Insulin	5 μg/ml
10	Dexamethasone	10- ⁸ M
	Ascorbic Acid	50 μg/ml
	Transferrin	$20-50~\mu g/ml$
	HSA	1%
	Linoleic Acid	6 μΜ
15	β-mercaptoethanol	5x10 ⁻⁵ M

Example 2

Bone marrow sample harvested from the iliac crest of the patient was washed twice with PBS. The nucleated cells were counted using methyl violet and plated at 5x10⁶ cells as unfractionated marrow per 10 cm tissue culture dish. For selection and expansion, the cells were maintained in Coon's modified Ham's F12 (F12) supplemented either with 10% FCS and 1 ng/ml FGF-2 (control culture) or the following defined components: EGF, PDGFbb, FGF-2, LIF, SCF, IGF-I, ascorbic acid, cholesterol, HSA, β-mercaptoethanol, dexamethasone, human holo- and apo-transferrin, selenium, biotin and sodium pantotenate. Table 2 below shows the preferred amounts of each component. To favor adhesion of the MSCs, the cells were first plated for 48 hours in F12 medium supplemented with 10% human serum and 1 ng/ml FGF-2. Thereafter, the medium was removed and the cells were extensively washed with PBS, and left for an additional 24-48 hours in F12 medium without any

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supplement. The defined mixture of factors was then added to promote cell proliferation.

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FGF-2 alone was the most active factor for maintenance of osteochondrogenic potential in mesenchymal stem cells. Selenium, biotin and sodium pantotenate were preferably included for cell viability. LIF and SCF were seen to improve the extent of cell proliferation, in particular in combination with IGF-1.

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Table 2

Medium Supplement for Serum Free Expansion of MSCs

10	INGREDIENT	CONCENTRATION
	(Basal medium: Coon's modified Ham's F	12)
	Human serum albumin	1-2%
	Transferrin (apo/holo)	$20-50~\mu\text{g/ml}$
	Ascorbic Acid	50 μg/ml
15	β-mercaptoethanol	$5 \times 10^{-5} M$
	Cholesterol	30 μg/ml
	Selenium	30 nM
	Biotin	33 µm
	Na pantotenate	17 μΜ
20	EGF/PDGF/FGF-2	1-10 ng/ml
	Dexamethasone	$10^{-8}M$
	IGF-I	5 ng/ml
	LIF .	5 ng/ml
	SCF	5 ng/ml

Studies have shown that PDGFbb by itself increases the osteogenic potential of MSCs when included in the phase of proliferation. This effect was found to be amplified by combining PDGFbb with FGF-2.

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Example 3

Growth Kinetics of Chondrocytes

At day 0, 5 x 10³ first passage cells were plated in each well of a 24-well plate in the presence of FCS. Upon adhesion, the FCS was removed, and the cells were extensively washed with PBS and left for 2-3 days in F12 without supplement to exhaust residual traces of serum. Proliferation was then reinduced by adding either 10% FCS or the mixture of defined components established for chondrocytes. Cell number was evaluated at different days via Thiazolyl blue (MTT) staining. Briefly, culture medium was removed and replaced with 0.5 ml of medium without supplement; then 25 µl MTT (Sigma, St. Louis, MO) stock solution (5 mg/ml) was added to each culture being assayed. After a 3 hour incubation the medium was removed and the converted dye solubilized with absolute ethanol. Absorbance of converted dye was measured at a wavelength of 570 nm with background subtraction at 670 nm.

The data obtained (see Figure 1) clearly show that the defined medium induces the chondrocytes to proliferate to a rate and extent comparable to those obtained in the presence of FCS.

Example 4

The differentiation potential of the chondrocytes expanded in serum free conditions was tested both *in vitro* and *in vivo*. For *in vitro* assay, the expanded cells were transferred in anchorage-independent conditions and maintained as a pellet culture for 2-4 weeks in the serum free medium previously shown by Johnstone et al. (Johnstone, B., Hering, T.M., Caplan, A.I., Goldberg, V.M. and Yoo, J.U. Exp. Cell Res. 238, 265-272, 1998) to induce chondrogenesis of serum expanded MSCs.

For in vivo assay, the expanded cells were implanted for 2 to 8 weeks in athymic mice either as a dense cell suspension or after embeddment in fibrin

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gel (Tissucol). At the term of the assays, the samples were fixed in formalin, Serial sections were processed for embedded in paraffin and sectioned. analysis and blue alcian blue) (toluidine and histological immunohistochemistry with collagen-specific antibodies.

Results indicated that, at variance with chondrocytes expanded in the presence of FCS, the chondrocytes expanded in serum free conditions directly reformed a cartilaginous structure both in vitro and in vivo, which stained metachromatic for toluidine blue, positive for alcian blue and type II collagen, and mostly negative for type I collagen. In contrast, in the case of the expansion in FCS, a total absence of full chondrogensis was observed both in vitro and in vivo; at most, a faint metachromatic staining was detected in some pellet cultures, but they always lacked well defined lacunae and well organized extracellular matrix.

These data illustrate a major advantage of the serum free system that allows chondrogenesis without the requirement of additional culturing in the presence of TGF- β 1 or other factors (Johnstone's inducing conditions). This may be due to the fact that chondrocytes, in nature, are not in contact with serum which may contain elements that inhibit chondrogenesis.

Example 5

The osteogenic potential of MSCs after expansion under serum free

defined conditions was tested in vivo by implantation of the expanded cells in athymic mice after adsorption on collagraft.

Several combinations of conditions were tested for bone formation in vivo. For all factor combinations, the medium contained Coon's modified Ham's F-12, dexamethasone, FGF-2, PDGFbb, EGF, transferrin, cholesterol, human serum albumin, biotin, selenium, Na pantotenate and ascorbic acid (concentrations as in Table 2). The combinations tested were 1) insulin; 2) IGF-1; 3) insulin and LIF; 4) insulin and SCF; 5) insulin, LIF and SCF; 6)

IGF-1 and LIF; 7) IGF-1 and SCF; and 8) IGF-1, LIF and SCF.

After 8 weeks of implantation, the samples were decalcified, included and processed for histology as above. The sections were stained with hematoxylin-eosin. All the conditions of expansion allowed the MSCs to reform bone tissue *in vivo*; however, the amount of bone formed varied from condition to condition. The combination of IGF-1, LIF and SCF provided an optimal expansion environment among the combinations tested.

CLAIMS

- 1. A serum free medium for cell culture, comprising a base minimum essential medium and
- 5 a) one or more growth factors; and
 - b) one or more sources of lipids and fatty acids.
 - 2. The serum free medium of claim 1, further comprising one or more steroids.
 - 3. The serum free medium of claim 1, further comprising
- 10 a) albumin;
 - b) an iron source;
 - c) one or more antioxidants;
 - d) a supplement for coenzyme transport in carboxyl group transfer reactions;
 - e) trace elements; and
- 15 f) vitamins.
 - 4. The serum free medium of claim 1 wherein the growth factor is selected from the group consisting of insulin, FGF-2, PDGFbb, EGF, LIF and SCF and IGF-1.
 - 5. The serum free medium of claim 2 wherein the steroid is dexamethasone.
- 20 6. The serum free medium of claim 1 wherein the lipid or fatty acid is cholesterol or linoleic acid.
 - 7. The serum free medium of claim 3 wherein the albumin is human serum albumin.
 - 8. The serum free medium of claim 3 wherein the iron source is transferrin.
- 9. The serum free medium of claim 8 wherein the iron source is human holoor apo-transferrin.
 - 10. The serum free medium of claim 3 wherein the antioxidant is β -mercaptoethanol or ascorbic acid.

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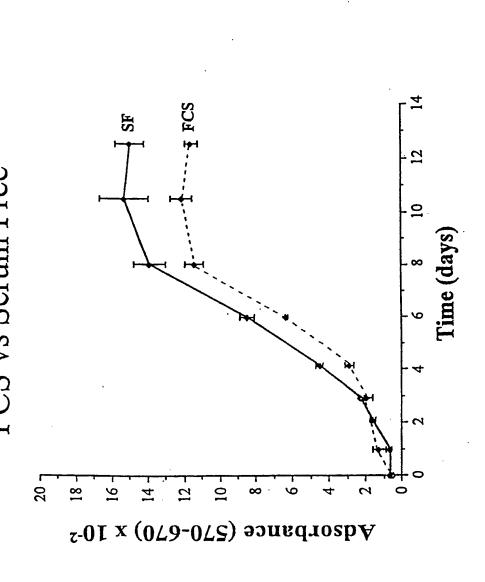
- 11. The serum free medium of claim 3 wherein the supplement for coenzyme transport in carboxyl group transfer reactions is biotin.
- 12. The serum free medium of claim 3 wherein the trace element is selenium.
- 13. The serum free medium of claim 3 wherein the vitamin is biotin or pantotenate.
- 14. A composition for the expansion of chondrocytes, comprising FGF-2, a fatty acid source, ascorbic acid, dexamethasone and insulin.
- 15. A composition for the expansion of chondrocytes, comprising a minimum essential medium and EGF, PDGFbb and FGF-2, ascorbic acid, linoleic acid, human serum albumin (HSA), β -mercaptoethanol, dexamethasone, insulin, human holo- and apo-transferrin.
- 16. A composition for the maintenance of mesenchymal stem cells, comprising selenium, biotin, sodium pantotenate, LIF, SCF and 1GF-1.
- 17. A composition for the maintenance of mesenchymal stem cells comprising a minimum essential medium and EGF, PDGFbb, FGF-2, LIF, SCF, IGF-I, ascorbic acid, cholesterol, HSA, β-mercaptoethanol, dexamethasone, human holo- and apo-transferrin, selenium, biotin and sodium pantotenate.

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FIG

hArticular Chondrocyte Growth Kinetics: FCS vs Serum Free



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Intern. Inal Application No PCT/EP 99/08482 CLASSIFICATION OF SUBJECT MATTER PC 7 C12N5/00 IPC 7 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) IPC 7 C12N Documentation searched other than minimum documentation to the extent that such documents are included. In the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. Category * WO 97 33978 A (LIFE TECHNOLOGIES INC) 1-13 X 18 September 1997 (1997-09-18) page 6, line 6 -page 7, line 3 page 8, line 18 -page 9, line 14 page 18, line 7 -page 19, line 6 tables 1-3 example 2 claims 1-4,6-8,2729,31,32,40,46-48 1-11,13WO 96 39487 A (OSIRIS THERAPEUTICS INC) X 12 December 1996 (1996-12-12) page 3, line 12 -page 4, line 14 page 5, line 19 -page 14, line 11 page 15, line 31 -page 16, line 14 page 21, line 7 - line 9 claims 1-31 -/--X Further documents are listed in the continuation of box C. X Patent family members are listed in annex. *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention Special categories of cited documents: *A* document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the International "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone filing date document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention connent or particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-ments, such combination being obvious to a person skilled in the art. "O" document referring to an oral disclosure, use, exhibition or "P" document published prior to the international filing date but "&" document member of the same patent family later than the priority date claimed Date of the actual completion of the international search Date of mailing of the international search report 20/04/2000 14 April 2000 **Authorized officer** Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl.

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